Author Queries

JOURNAL: LCSS MANUSCRIPT: 162339

- **Q1** Please define NADPH.
- Q2 Please spell out NSW.
- **Q3** Please verify journal title-word missing?
- Q4 Word missing in journal title?
- **Q5** Please spell out publishers.

Communications in Soil Science and Plant Analysis, 37: 1-11, 2006 Copyright © Taylor & Francis Group, LLC ISSN 0010-3624 print/1532-2416 online 1 DOI: 10.1080/00103620600623590 2 3 4 5 6 7 8 Kikuyu Grass: A Valuable Salt-Tolerant 9 Fodder Grass 10 11 12 Mathangi Radhakrishnan, Yoav Waisel, 13 and Marcelo Sternberg 14 Department of Plant Sciences, Tel Aviv University, Tel Aviv, Israel 15 16 17 18 Abstract: The turf grass Pennisetum clandestinum Hochst. (kikuyu grass) is one of the 19 candidate plants for utilization and reclamation of salinized areas. The capability of 20 kikuyu grass to grow under saline conditions was tested during 6 months, under 21 various irrigation treatments (tap water control, 80-mM, 150-mM, 200-mM, 240-22 mM NaCl). Plant biomass production was visibly affected only at NaCl concentrations 23 greater than 150-mM NaCl. Plant growth and plant regeneration capability in the 200-24 and 240-mM NaCl treatments gradually decreased as the experiment progressed in 25 time. The photosynthetic potential of the plants remained unchanged and was neither affected by the treatment nor with time. Proline content of leaves as well as the 26 content of Na⁺ and Cl⁻ increased with increasing salinity stress. Apparently, kikuyu 27 grass can withstand moderate concentrations of NaCl for prolonged periods and 28 under repeated mowing. Thus, this grass can be used as a potential ground cover and 29 as fodder grass in saline habitats. 30 31 Keywords: Pennisetum clandestinum, proline, salinity, sodium chloride 32 33 34 **INTRODUCTION** 35 36 Salt stress is one of the most serious environmental factors limiting the pro-37 ductivity of crop plants (Ashraf 1999). Saline soils present special challenges 38 for plant subsistence and growth. Moreover, salinity plays a major role in 39 natural ecosystem functioning, limiting plant development, particularly in 40 arid and semiarid regions. In such ecosystems, reduced precipitation leads 41 42 43 Received 30 August 2005, Accepted 6 January 2006 44 Address correspondence to Yoav Waisel, Department of Plant Sciences, Tel Aviv

45 University, Britannia Building, Tel Aviv, Israel. E-mail: yoavw@tauex.tau.ac.il

M. Radhakrishnan, Y. Waisel, and M. Sternberg

to reduced leaching of salt from the soil and consequently to salinization of the 46 upper soil horizon. Such a process leads to the alteration of the soil structure, 47 causing severe soil erosion, plant cover reduction, and a deterioration of the 48 49 natural vegetation. Amelioration of saline soils can be achieved by the introduction of salt-resistant ground cover species (Chapman 1960). Certain 50 varieties of kikuyu grass (Pennisetum clandestinum Hochst.) are known to 51 be tolerant to salinity (Russell 1976; Skerman and Riveros 1990), tolerant 52 to drought (Whiteman 1990), and tolerant to water logging (Dale and Read 53 1975). Thus, this species seems to be a good candidate for planting and use 54 in such habitats. 55

Kikuyu grass is a perennial grass native to East and Central Africa 56 (Skerman and Riveros 1990) that was introduced to many parts of the world 57 (Rumball 1991; Herreroborgonon, Cristobal, and Crespo 1995). Considering 58 its fast growth, dense ground cover, and well-developed root system, kikuvu 59 grass can be an exceptional species for erosion control on desert edges and 60 salinized soils. Kikuyu grass can also be used as a pasture plant, because of 61 its good nutritive properties (Butler and Bailey 1973; Marais, Figenschou, 62 and de Figueredo 1992). 63

Moreover, its capability to regenerate rapidly following repeated mowing is a highly important trait of fodder plants, especially for those that are grown under saline conditions (Gugenheim and Waisel 1977).

Salt tolerance of plants involves the preservation of a basic ionic 67 balance in their cells and certain metabolic changes that decrease salt 68 injury. Accumulation of Cl⁻ was reported as one of the means that 69 enables salt tolerance in some plants (Azmi and Alam 1990; Ashraf and 70 O'Leary 1995). Increased Na⁺ content generally disturbs the nutrient 71 balance and osmotic regulation of sensitive plants but is controlled in salt-72 tolerant plants, where it plays an important role in their adaptation 73 (Waisel 1989). 74

75 The decline in productivity of many salt-affected plant species that are subjected to excess salinity is often associated with a reduction in photosyn-76 thetic capacity (Long and Baker 1986). One of the common adaptive 77 responses of plants to salt stress is expressed by their proline metabolism. 78 Proline is an important factor in establishment of an osmotic equilibrium in 79 salt-affected plants, and thus is a good method for monitoring of stress 80 tolerance of such plants (Delauney and Verma 1993; Sidari, Panuccio, and 81 Muscolo 2004). The fast and aggressive reproduction and spread of kikuyu 82 grass made it a weed in numerous agricultural and recreational areas (Wilen 83 et al. 1995; Wilen and Holt 1996). However, such characteristics could be 84 an advantage for reclamation of salt-affected sites. Thus, the present study 85 has tackled the following questions: 86

87 88

89

a. How do increasing concentrations of NaCl affect above- and belowground biomass production of kikuyu grass?

b. Does salinity affect the photosynthetic efficiency of this plant species?

Kikuyu Grass

91	c.	How much is the accumulation of ions in leaves of kikuyu grass affected
92		by the increasing salt concentrations of the irrigation water?
93	d.	What is the role of proline in osmotic adaptation of this species?

- 94
- 95
- 96

MATERIALS AND METHODS

97 98 99

Plant Materials and Growth Conditions

Cuttings and runners of P. clandestinum were grown in pots filled with 10 kg 100 of sand. The pots were watered with a nutrient solution equivalent to half of 101 Hoagland's nutrient solution. The plants were grown in a ventilated green-102 house ($\sim 25^{\circ}$ C) under long-day conditions (16/8-h photoperiod). 103

104 105

106

Experimental Treatments

107 The plants were irrigated daily with different NaCl treatments (80-mM, 108 150-mM, 200-mM, and 240-mM of NaCl, supplemented with the nutrient 109 solution), with 12 replicates per each treatment. Control pots were treated 110 with nutrient solution only. The treatments were continued for 6 months. 111

112

113 **Plant Biomass**

114

115 Plants were pruned regularly every month at the height of 4 cm above the 116 ground. The clipped shoots were oven dried at 80°C for 2 days and then 117 weighed. At the end of the experiment, 80 days after transplantation, the 118 plants were removed from the pots. Shoots and roots were separated, 119 washed, and dried at 80°C for 2 days, and the dry weight was then determined.

120 121

123

Photosynthetic Yield Measurements 122

Pulse amplitude modulated fluorometry (PAM, Walz Gmbh, Effeltrich, Germany) 124 was used to measure the photosynthetic quantum yield of dark adapted fully 125 developed leaves of the treated plants after 6 months of treatment. Leaf clips 126 (Dark Leaf Clip DLC-8) were placed on different leaves, taken randomly from 127 the treated plants, to have them completely dark adapted for a period of 30 min. 128 Maximal quantum yields were evaluated using the equation 129

$$\frac{F_v}{F_m} = \frac{(F_m - F_o)}{F_m}$$

132

where F_{ν} is the variable fluorescence, F_{ρ} is the fluorescence measured after dark 133 adaptation, and F_m is the maximal fluorescence of the dark-adapted leaves after 134 the application of a 0.8-s pulse of light that is saturating for photosynthesis 135

M. Radhakrishnan, Y. Waisel, and M. Sternberg

136 ($\sim 6000 \,\mu\text{mol}$ photons m⁻² s⁻¹) (Beer et al. 1998; Beer et al. 2000; Kitajima and 137 Butler 1975).

- 138
- 139

140 **Proline Estimation**

141

Fresh leaf segments, taken from plants of the different treatments, were cut and ground in liquid nitrogen. Approximately 2 mL of 3% sulfasalicylic acid were added, and the test tube was then centrifuged at 14 k rpm for 5 min. Proline was determined using ninhydrin acid reagent according to Bates, Waldren, and Teare (1973) and L-proline as standard.

147

¹⁴⁸ Determination of Na⁺ and Cl⁻

Dried leaves, taken from each of the different treatments, were weighed and
ashed in the furnace at 550°C for 6 h. The ash was then dissolved in 5 mL
of 2 M nitric acid, diluted, and measured by flame spectrophotometry.
Chloride was determined with a chloride analyzer (MKII Chloride Analyzer
926, Sherwood Scientific Ltd., Cambridge, England).

155 156

157 Statistical Analysis

158

159 Data was analyzed using one- and two-way analyses of variance (ANOVA), 160 and treatment means of the significant differences were separated using the 161 Tukey test (p < 0.05).

- 162
- 163

¹⁶⁴ **RESULTS**

165

¹⁶⁶ Effects of Salinity on Biomass Production

167

Exposure of kikuyu grass to all the salinity treatments for 6 months reduced the
aboveground shoot biomass as well as the root growth (Figure 1). Highest inhibition was noted at the 200-mM NaCl treatment. Root biomass was strongly
affected by the salt treatment, and differences in root growth were found (cf.
Shalhevet, Huck, and Schroeder 1995). The root-shoot ratio dropped down in
the salt-exposed plants and was lowest in the 240-mM NaCl treatment (Figure 2).

175

176 Effect of Shoot Pruning at Definite Intervals

177

178 Shoot regeneration was affected by all irrigation treatments with saline water.

179 Inhibition was positively correlated with the salinity treatments. For example,

180 shoot production for the 200-mM treatment at the first clipping was only 47%

Kikuyu Grass



Figure 1. Effects of increasing NaCl concentrations on dry root and shoot biomass of kikuyu grass (mean \pm SE). Means with common letter do not differ from each other (Tukey HSD, p < 0.001). Capital letters refer to shoot grouping and lowercase letters indicate root grouping.

of the production of the control plants but 74% for the plants of the 240-mM
 treatment.

After 6 months, the regrowth had dropped down to 88% of the initial values at 200 mM of NaCl and was practically abolished at the 240-mM treatment, leading eventually to the death of many of the plants of this treatment (Figure 3).



220 221 222

203

210 211

212

213

214

215

216

217

218

219

Figure 2. Effect of increasing NaCl concentration on root-shoot ratio of kikuyu grass (mean \pm SE). Means with common letters, do not differ from each other (Tukey HSD, p < 0.05).

5



Figure 3. Effect of salinity on dry weight production: pruning at different times.

Proline Accumulation

To clarify the mechanisms of osmotic adaptation by kikuyu grass plants, proline content of the leaves of the treated plant was monitored (Figure 4). Up to a four-fold increase in proline content (15.14 μ mol/g fresh weight) was measured in salt-stressed shoots of the 240-mM NaCl treatment as compared to 3.5 μ mol/g fresh weight in the 80-mM NaCl treatment.



Figure 4. Effects of salt stress on proline accumulation (μ mol g⁻¹ FW) of leaves in kikuyu grass exposed to different salt concentrations (mean \pm SE). Means with common letters do not differ from each other (Tukey HSD, p < 0.001).

3/13/2006

Kikuyu Grass

271 272

Photosynthetic Response of Chlorophyll Fluorescence to Salt Stress

The photosynthetic quantum yield of leaves of the treated plants was measured. The photosynthetic yield obtained was similar for all treatments, except for the 200-mM NaCl treatment (-70%). Apparently the applied salt stress had no substantial effect on PSII photochemistry (Figure 5).

277 278 279

280

Ion Content in Leaves

Sodium and chloride concentrations of the leaves of the treated kikuyu grass 281 plants increased with the salinity of the medium. Sodium and chloride content 282 in the control were 9.5 mg g^{-1} and 21.5 mg g^{-1} respectively and reached 17.7 mg g^{-1} and 30.72 mg g^{-1} respectively in the 200-mM treatment. 283 284 Leaves of plants with high ion accumulation exhibited bleaching and 285 scorched leaves that eventually lead to death. In some plants of the 240-286 mM NaCl treatment, where leaf turnover was fast, sodium and chloride 287 content in the leaves was rather low (Figure 6), apparently representing the 288 ion content only of the young regenerating leaves. 289

290 291

292 293

300 301

302

303

304

305

306

307

308

309

310

DISCUSSION

Growth of kikuyu grass plants was little affected by low salt concentrations of the irrigation water. The plants showed a gradual decrease in leaf production that was distinct from the 150-mM treatment and up. This confirmed the results that were previously reported (Skerman and Riveros 1990; Muscolo, Panuccio, and Sidari 2003) that marked the salt tolerance of this grass at the 200-mM NaCl concentration.





Figure 5. Maximal photosynthetic quantum yield (F_v/F_m) in leaves of kikuyu grass exposed to different concentrations of salt (mean \pm SE). Means with common letters do not differ from each other (Tukey HSD, p < 0.05).

7

318

319

320

321

322

323

324

325

326

327



Figure 6. Effect of salinity on ion content in leaves of kikuyu grass exposed to different concentrations of salt stress (mean \pm SE). Means with common letters do not differ from each other (Tukey HSD, p < 0.001).

331 332

Growth inhibition under saline conditions might be caused either by the 333 lack of osmotic adaptation or by specific poisoning. The lack of osmotic 334 adjustment reduces water uptake and causes physiological drought. This has 335 long been considered the major cause of salinity injury to plants (Waisel 336 1972; Levitt 1980). Adaptation is reached either by accumulation of 337 inorganic ions or by accumulation of compatible solutes. Some grasses are 338 capable of reaching a fast and complete adjustment under salinity stress 339 340 (Marcum and Murdoch 1990). Judging from the changes in proline content of the salt-exposed kikuyu plants, such a trait also applies to them. 341

In the present study, we also observed the inhibitory effects of repetitive leaf pruning. Such a trait lowers the value of plants planned to be used as turf grasses or grown for grazing. However, at moderate salinities, even a reduced production is of value.

Could it be that reduced growth resulted from a reduction in photosyn-346 thesis? Several reports have mentioned that salt stress is enhancing photoinhi-347 bition (Sharma and Hall 1991) and by that affect net photosynthesis. However, 348 the presented results imply that the photosynthetic potential of kikuyu grass 349 was not really lowered by the salinity treatments. Because the stolons and 350 rhizomes were not damaged by the repeated leaf clippings, we have to 351 conclude that the regeneration potential of the plants was lowered by the 352 removal of photosynthesizing leaves and not by reduction of PSII yield. Con-353 comitantly, adaptation of the PSII system in salt-stressed leaves can be 354 explained as an important strategy of plant adaptation particularly in arid 355 and warm regions (Lu and Zhang 1998). Another cause for the decline in 356 growth under saline conditions appears to be related to an excessive buildup 357 of ions in leaf tissues (Muscolo, Panuccio, and Sidari 2003) and to the 358 inability of plants to produce new leaves to replace the senescent ones (cf. 359 Guggenheim and Waisel 1977). This was reconfirmed in the present study. 360

Kikuyu Grass

Accumulation of sodium and chloride ions is instrumental for leaf 361 osmotic adjustment in halophytes (Waisel 1989; Marcum 1999) and is 362 enhanced by salinity (Rawson, Long, and Munns 1988; Ghoulam, Foursy, 363 and Fares 2002). Such adaptation is limited to a certain concentration range. 364 In the present study, it has peaked in plants of the 200-mM treatment. The 365 decline in the average leaf content of sodium and chloride, at high concen-366 trations of external NaCl (cf. Hussain, Caemerer, and Munns 2004) is not 367 because of the reduction in ion accumulation by the individual leaves but 368 because of an unbalanced leaf turnover, a fast shedding of the salt-loaded 369 old leaves concomitantly with some production of low-salt young leaves. 370

Proline accumulation is often considered to be a major factor involved in 371 osmotic adaptation and is used as a measure for stress tolerance (Delauney and 372 Verma 1993). Accumulated proline in plants acts not only as an osmolyte. It 373 involves an improved NADPH supply and is an "easy to handle" energy 374 01 reservoir as well as an energy shuttle between plastids and mitochondria 375 (Hare and Cress 1997). Thus, accumulation of proline serves a multitude of 376 adaptation systems. Nevertheless, its role in plant adaptation to salinity 377 remains controversial. Thus, the osmotic adaptation of kikuyu grass plants 378 is achieved by two means: sodium and chloride accumulation on one hand 379 and proline accumulation on the other. 380

Findings of this study present kikuyu grass as a reasonably good turf and forage grass, as well as a cover grass that can endure relatively high soil salinity. Kikuyu grass can be a good candidate to combat the ongoing land degradation and can play an important role in saline land reclamation, combating the spread of salinization in arid and semiarid regions.

386

387 388 389

ACKNOWLEDGMENTS

We thank our colleagues and the members of our laboratories for their help
and support. This research work was performed as a part of postdoctoral
study by M. Radhakrishnan supported by the Israel Council of Higher
Education and the Israeli Ministry of Foreign Affairs.

- 395
- 396

398

397 **REFERENCES**

- Ashraf, M. (1999) Breeding for salinity tolerance proteins in plants. *Crtical Review Plant Sciences*, 13: 17–42.
- Ashraf, M. and O'Leary, J.W. (1995) Distribution of cations in leaves of salt-tolerant and salt-sensitive lines of sunflower under saline conditions. *Journal of Plant Nutrition*, 18 (12): 1811–2375.
- Azmi, A.R. and Alam, S.M. (1990) Effect of salt stress on germination growth leaf
 anatomy and mineral element composition of wheat cultivars. *Acta Physiologia Plantarum*, 12: 215.

9

Bates, L.S., Waldren, R.P., and Teare, I.D. (1973) Rapid determination of free proline

M. Radhakrishnan, Y. Waisel, and M. Sternberg

407	for water-stress studies. Plant and Soil, 39: 205-207.	
408	Beer, S., Larsson, C., Poryan, O., and Axelsson, L. (2000) Photosynthetic rates of Ulva	
409	(Chlorophyta) measured by pulse amplitude modulated (PAM) fluorometry.	
410	European Journal of Phycology, 35: 69–74.	
411	Beer, S., Vilenkin, B., Weil, A., Veste, M., Susel, L., and Eshel, A. (1998) Measuring	
411	photosynthetic rates in sea grasses by pulse amplitude modulated (PAM) fluorome-	
412	try. Marine Ecology Progress Series, 174: 293–300.	
413	Butler, G.W. and Bailey, R.W. (1973) Chemistry and Biochemistry of Herbage;	
414	Academic Press: London.	
415	Chapman, V.J. (1960) Salt Marshes and Salt Deserts of the World; Leonard Hill:	
416	London.	
417	Dale, A.B. and Read, J.W (1975) Irrigation of kikuyu pastures, Kikuyu Research	
418	Report; NSW Department of Agriculture, Government of NSW: Victoria, Australia.	Q2
419	Delauney, E.F. and Verma, D.P.S. (1993) Proline biosynthesis and osmoregulation in	
420	plants. Plant Journal, 4: 215–223.	
421	Gnoulam, C., Foursy, A., and Fares, K. (2002) Effect of sait stress on growin, inorganic	
422	cultivers Environmental and Experimental Rotany 47:30,50	
423	Guagenheim I and Waisel V (1077) Effects of salinity temperature and nitrogen fer.	
424	tilization on growth and composition of Bhodes grass (Chloris gavang Kunth), Plant	
425	and Soil 47: 431-440	
426	Hare PD and Cress WA (1997) Metabolic implications of stress-induced proline	
427	accumulation in plants. <i>Plant Growth Regulation</i> , 21: 79–102.	
428	Herreroborgonon, J.J., Cristobal, J.C., and Crespo, M.B. (1995) <i>Pennisetum clandesti</i> -	
429	num Hochst Ex Chiov Poaceae: An African grass in Europe. Israel Journal Plant	
430	<i>Sciences</i> , 43: 159–162.	
431	Hussain, S., Caemerer, S.V., and Munns, R. (2004) Control of salt transport from roots	
422	to shoots of wheat in saline soils. Functional Plant Biology, 31: 1115-1126.	
432	Kitajima, M. and Butler, W.L. (1975) Quenching of chlorophyll fluorescence and	
435	primary photochemistry in chloroplasts by dibromothymoquinone. Biochimica et	
425	Biophysica Acta, 376: 105–115.	
455	Levitt, J. (1980) Responses of Plants to Environmental Stresses; Academic Press:	
430	New York.	
437	Long, S.P. and Baker, N.R. (1986) Saline terrestrial environments. In <i>Photosynthesis in</i>	
438	<i>Contrasting Environments</i> ; Baker, N.K. and Long, S.P. (eds.); Elsevier: New York,	
439	05-110.	
440	stressed sorghum Australian Journal of Plant Physiology 25: 317–324	
441	Marais IP Figenschou DI and de Figueredo M (1992) Effect of nutrient calcium	
442	on the cellwall composition and digestibility of Kikuyu grass <i>Pennisetum clandesti</i> -	
443	num Hochst, Journal Grassland Society of South Africa, 9: 2–75.	02
444	Marcum, K.B. (1999) Salinity tolerance mechanisms of grasses in the subfamily Chlor-	QS
445	idoideae. Crop Science, 39: 1153–1160.	
446	Marcum, K.B. and Murdoch, C.L. (1990) Growth responses ion relations and osmotic	
447	adaptations of eleven C4 turfgrasses to salinity. Agronomy Journal, 82: 892-896.	
448	Muscolo, A., Panuccio, M.R., and Sidari, M. (2003) Effects of salinity on growth	
449	carbohydrate metabolism and nutritive properties of Kikuyu grass Pennisetum clan-	
450	destinum Hochst. Plant Science, 164: 1103-1110.	

Kikuyu Grass

451 452	Rawson, H.M., Long, M.J., and Munns, R. (1988) Growth and development in NaCl ⁻ - treated plants, I: Leaf Na ⁺ and Cl ⁻ concentrations do not determine gas exchange of leaf blades in barlay. <i>Australian Journal of Plant Physiology</i> , 15: 519–527	
453 454	Rumball, P.J. (1991) The performance of several sub-tropical grasses in the northland and bill postures. New Zeeland Lournal of Activity 24, 275–282	
455	Russell J.S. (1976) Comparative salt tolerance of some tropical and temperate legumes	
456	and tropical grasses. Australian Journal Experimental Agriculture, 16: 103–109.	04
457	Shalhevet, J., Huck, M.G., and Schroeder, B.P. (1995) Root and shoot growth	×.
458	responses to salinity in maize and soybean. <i>Agronomy Journal</i> , 87: 512–516.	
459	Sharma, P.K. and Hall, D.O. (1991) Interaction of salt stress and photoinhibition on photosynthesis in barley and sorghum <i>Journal Plant Physiology</i> 113: 614–619	~
460	Sidari, M., Panuccio, M.R., and Muscolo, A. (2004) Influence of acidity on growth and	Q4
461	biochemistry of Pennisetum clandestinum. Biologia Plantarum, 481: 133-136.	
462	Skerman, P.J. and Riveros, F. (1990) FAO Plant Production and Protection Series;	Q5
463	FAO & UN: Rome, Italy.	
404	Waisel, Y. (1972) Biology of Halophytes; Academic Press: New York. Waisel, Y. (1989) Screening for salt resistance. In <i>Proceedings of 21st Colloquium of</i>	
405	the International Potash Institute: Methods of K-Research in Plants; International	
467	Potash Institute: Bern Switzerland.	
468	Whiteman, P.C. (1990) Tropical Pasture Science; Oxford University Press: Oxford,	
469	UK. Wilen C.A. Halt J.S. Ellatoond N.C. and Show D.C. (1005) Construin diversity of	
470	kikuvu grass <i>Pennisetum clandestinum</i> populations in California <i>Weed Science</i> 43:	
471	209–214.	
472	Wilen, C.A. and Holt, J.S. (1996) Spatial growth of Kikuyu grass Pennisetum clandes-	
473	tinum. Weed Science, 44: 323–330.	
474		
475		
476		
477		
478		
479		
480		
481		
482		
483		
404		
485		
487		
488		
489		
490		
491		
492		
493		
494		
495		